

## Effect of temperature on fatty acid profile of *Nostoc spongiaeforme* (freshwater) and marine water *Nostoc calcicola* (marine water): A comparative study

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The Fatty acid profiling in *Nostoc spongiaeforme* and *Nostoc calcicola* was undertaken at varying growth temperature. The extraction of fatty acid was done by chloroform: methanol method and identification of various SFA, MUFA and PUFA was performed using GC-MS. The result of our study showed that amount of fatty acid content varied with varying temperature. At low temperature, the content of unsaturated fatty acid was maximum in both the *Nostoc* species but the amount of saturated fatty acid was reduced under the same condition. Whereas, at extreme temperature unsaturated fatty acid was least in both *Nostoc* species and saturated fatty acid was maximum. Both species showed higher amount of fatty acid content at 20°C and 30°C. Our study concluded that the best temperature for mesophilic cyanobacteria to produce higher amount of fatty acid content is in the range of 20-30°C.

*Key words:* Cyanobacteria; Fatty acids; GC-MS; *Nostoc*, Temperature

*Abbreviations:* SFA, Saturated fatty acids; MUFA, Monounsaturated fatty acids; PUFA, Polyunsaturated fatty acids; GC-MS, Gas chromatography- Mass spectroscopy

Rapid industrialization and population growth have put pressure on finite energy sources (Petroleum-derived fuel) (Shafiee and Topal, 2009). Due to this, the emission of greenhouse gases has also intensified and hence causing climate change and hampering the biosphere (Chisti, 2007). Due to the above mentioned reasons, there is a need to call for alternative sustainable and renewable energy sources (Demirbas and Demirbas, 2011). Production of biofuel from different biomass could be an important alternative and sustainable step to fossil-derived fuels (Li et al., 2011). Biofuels are characterized into three generations based on the type of feedstock used. The first-generation biofuels are derived from different crops like corn, soybean, sunflower, palm oil, and rapeseed. *Jatropha*, *Miscanthus*, *Switchgrass* and other organic wastes are used to derive second-generation biofuel. As microalgae and cyanobacteria does not require agricultural or arable land for production and has high photosynthetic efficiencies, biomass productivities and high lipid content. They are considered as the third and most reliable material for biofuel production (Hariskos and Posten, 2014; Tredici, 2010). Moreover, in microalgae storage lipids are in the form of neutral lipids or triacylglycerols, which can be esterified to fatty acid methyl esters (FEMs), most suitable for biofuel production (Ge et al., 2017). Moreover, cyanobacteria have a great adaptation ability to various environmental conditions and easy to cultivate.

Among the different abiotic factors, temperature is one of the vital parameters which influence the overall growth of microalgae and cyanobacteria. Temperature is also responsible for directly influencing the lipid content and fatty acid composition of algae and cyanobacteria but the nature and magnitude of temperature effect may vary species to species (Guschina and Harwood, 2006). In order to thrive the change in the ambient temperature, organisms alter their fatty acid composition which is considered as a mechanism to maintain normal cellular function (Thompson, 1996). The decrease in the growth temperature increases the ratio of unsaturated to saturated fatty acids (Renaud et al., 2002). In order to sustain low temperature and to maintain greater

membrane fluidity at low temperature, organisms often increase the ratio of unsaturated fatty acid over saturated fatty acid whereas, at high temperature, the organisms cell membrane becomes more saturated to attain higher rigidity (Hilditch and Williams, 1964). For instance, the tobacco plant was made tolerance to cold by introducing the omega-3 fatty acid desaturase gene (*fad7*), which increased the trienoic fatty acid (16:3 and 18:3) content. At the same time, suppression of *fad7* has resulted in a decrease in the trienoic fatty acid which helped the tobacco plant to get acclimatize to high temperatures (Murakami et al., 2000). However, it is vital to note that the response of growth temperature on the organism is species-specific, with no overall consistent relationship between fatty acid unsaturation and temperature (Guschina and Harwood, 2006).

Mesophilic cyanobacteria cultures grow within a high diurnal temperature range of 25-30°C throughout the year. We are curious to study the effect of different growth temperature on the fatty acid composition of mesophilic nitrogen-fixing cyanobacteria from two different habitats. The purpose of our work was to explore the effect of different growth temperatures, within the range of 10°C-40°C, on the fatty acid composition of two *Nostoc* species i.e. *Nostoc spongiaeforme* (Freshwater) and *Nostoc calcicola* (Marine water) nitrogen-fixing cyanobacteria.

## MATERIALS AND METHODS

### Culture of *N. spongiaeforme* and *N. calcicola*

*N. spongiaeforme* Agardh, (a freshwater cyanobacterium present in rice fields) culture was regularly grown in the aseptic BG-11 culture medium. *N. calcicola* Brébisson ex. Bornet & Flahault, (a marine cyanobacterium, procured from National Facility for Marine Cyanobacteria, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India) was grown in sterile ASN III culture medium, following Rippka et al., 1979, kept at optimum temperature (30°C) for the growth and was maintained under continuous fluorescent white light (80  $\mu\text{mol photon m}^{-2} \text{s}^{-1}$ ) with a 16:8 h photoperiod. The culture growth was monitored on alternate days by measuring absorbance at 680 nm (Wang et al., 2012) using a spectrophotometer (UV-2450, Shimadzu model,

Japan). The cells in the exponential growth phase (13<sup>th</sup> and 25<sup>th</sup> day) were harvested and lyophilized and stored at -20°C until required.

#### Temperature

Once the exponential growth phase was set, both *Nostoc* species were grown at different experimental temperatures (10, 20, 30, and 40 °C) on the 13<sup>th</sup> and 25<sup>th</sup> day in sterilized BG-11 and ASN III medium.

#### Fatty acid extraction

50 mg of lyophilized tissue was transferred to 50ml graduated centrifuge tube with 5ml of pre-cooled (4°C) 1:2 (v/v) chloroform: methanol and spin-vortex for 3 min and sonicated for 15min in a sonicator. Samples were centrifuged at 4°C, 10,000 x g for 10 min and supernatant was pooled in a fresh 50 ml tube. To the pellet, 2.5 ml pre-cooled 1:1 (v/v) chloroform: methanol was added, vortexed and centrifuged for 10min at 10,000 x g at 4°C and supernatant were pooled out and this step was repeated for 3-4 times. Pooled supernatant was filtered using Whatman filter paper and wash by mixing with an equal volume of potassium phosphate buffer (50 mM; pH-7.5). The filtrate was centrifuged for 10 mins at 10,000 x g at 4°C and the lower organic phase was collected and total lipids were dried under N<sub>2</sub> using a sample concentrate and total dried lipid was stored at -20°C for further analysis.

#### Conversion of total lipid to fatty acid methyl esters (FAME) by transmethylation

The fatty acid profile of *N. spongiaeforme* and *N. calcicola* were performed by Gas Chromatography coupled with Mass spectrophotometer. Total lipid dissolved in 1 ml of methanolic NaOH and kept for drying in the oven at 80°C for 5 hours. After drying, 2ml of 5% methanolic HCL was added and vortex for 20 sec. Tubes were kept for drying at 80°C for 5 hours. To this 1 ml of water (Milli Q) and 2 ml of 100% Hexane (HPLC grade) was added, spine and allow to settle down. Top layer was transferred into a new vial and this step was repeated 3 times. At the last stage, the upper layer was collected and 1 ml saturated sodium bicarbonate was added, spine and allow to settle down and the upper layer was collected into the new vial. To this, 1 ml sterile water was added with few crystals of CaCl<sub>2</sub> and allow for

settle down and again upper layer was transferred into the new vial and dry with N<sub>2</sub> and before injecting into the GC-MS injector, the sample was dissolved in 500 µl of 100% hexane.

#### GC-MS analysis

Chromatography was performed using a Shimadzu Mass spectrometer-2010 plus (CIF, SPPU, Pune) equipped with GC-MS-TQ8030, equipped with an auto-sampler (AOC-20s). An electron ionization system coupled with an ionization energy of 70eV was used for GC-MS detection. The carrier gas used was He at a flow rate of 1ml per minute. Column oven and injector temperatures were set 50°C and 270°C. The compound was identified by matching the mass spectra with the inbuilt library National Institute of Standard and Technology (NIST).

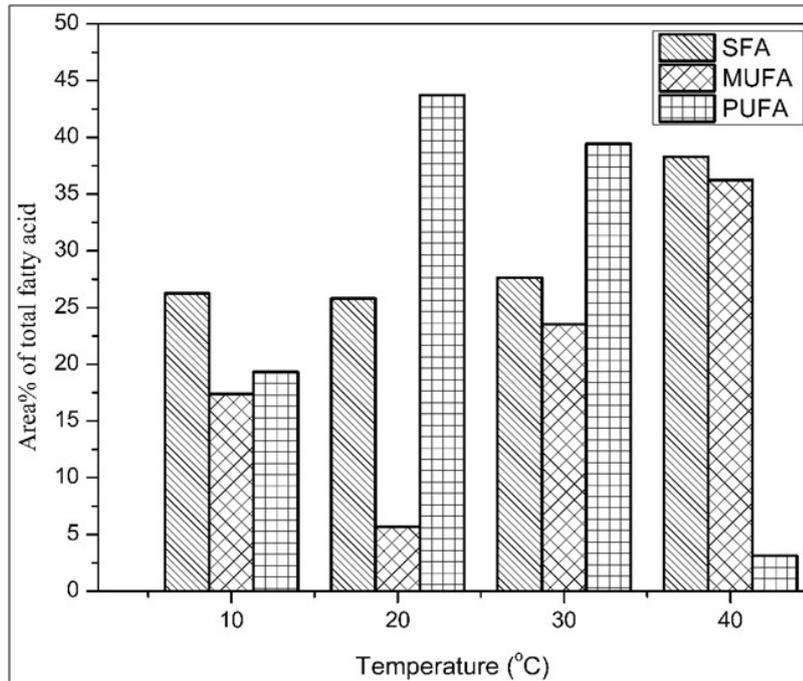
## RESULTS

Fatty acid methyl ester (FAME) composition of two *Nostoc* species at different temperatures is shown in (Fig.1, 2 and Table 1, 2). From our study, it was shown that the saturated and mono-unsaturated fatty acid increased by increasing the temperature in *N. spongiaeforme*. *N. spongiaeforme*, at 40°C showed the maximum percentage of saturated and mono-unsaturated fatty acid (38.28 and 36.24% respectively) as compared to control (30°C). Whereas, under the same conditions, the poly-unsaturated fatty acid was reduced to 3.14% compared to control. Through our study, it was also observed that *N. spongiaeforme* when grown at a relatively lower temperature (20°C), resulting in an increase in the poly-unsaturated fatty acid 43.71% as compared to control. Whereas, at 10°C only 19.33% poly-unsaturated fatty acid was observed which was far less than control (39.44%).

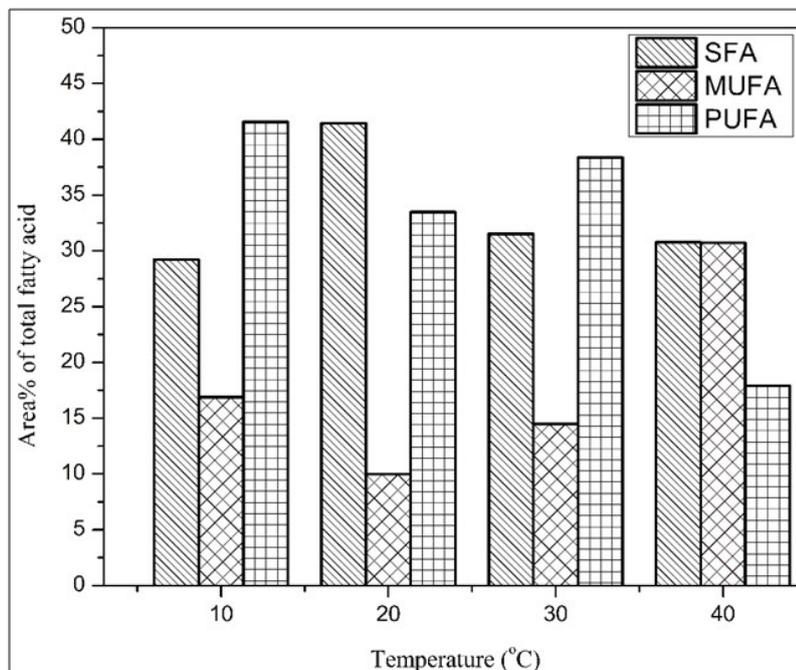
*N. calcicola* also showed variation in the fatty acid composition when grown at different growth temperatures. The percentage of saturated fatty at 40°C (30.77%) was more or less similar to control (31.52%), whereas mono-unsaturated fatty acid was increased to 30.73% compared to control (14.48%). Where, at same temperature, the poly-unsaturated fatty acid was reduced 17.91% compared to control (38.36%). Cultures grown at 10°C, resulted into reduce saturated and

monounsaturated fatty acid compared to higher temperature treatment, whereas poly-unsaturated fatty acid was increased to 41.45% as compared to control. On the other hand, 20°C showed highest amount of

saturated fatty acid and the least amount of monounsaturated fatty acid among all the growth temperatures whereas, 33.46% of poly-unsaturated fatty acid was detected.



**Figure 1:** Effect of different growth temperatures on the total methyl esters of fatty acids of *N. spongiaeforme*. **Abbreviations** - SFA: saturated fatty acid; MUFA: monounsaturated fatty acid; PUFA: polyunsaturated fatty acid. *N. spongiaeforme*: *Nostoc spongiaeforme*.



**Figure 2:** Effect of different growth temperatures on the total methyl esters of fatty acids of *N. calcicola*. **Abbreviations** - SFA: saturated fatty acid; MUFA: monounsaturated fatty acid; PUFA: polyunsaturated fatty acid; *N. calcicola*: *Nostoc calcicola*

**Table 1:** Analysis of Fatty acid profile based on the methyl ester of fatty acids (Area% of fatty acids) of *Nostoc*

Fatty acid	Percent of fatty acids (% of total fatty acids)			
	10°C	20°C	30°C	40°C
9-Hexadecenoic acid, methyl ester (C16:1)	3.02	ND	15.18	3.97
Hexadecenoic acid, methyl ester (C16:0)	13.92	21.55	23.94	17.38
9,12-Octadecadienoic acid, methyl ester (C18:2)	10.28	2.32	18.98	2.86
9,12,15-Octadecatrienoic acid, methyl ester	8.34	27.16	18.93	ND
9-Octadecenoic acid methyl ester (C18:1)	14.36	5.01	7.62	32.27
Octadecanoic acid (C18:0)	10.81	3.99	3.16	14.69
Glycidyl palmitate	1.52	ND	0.29	0.62
Ethyl 9,12-hexadecadienoate	0.71	1.95	ND	ND
Methyl tetradecanoate (C14:0)	ND	0.20	0.23	ND
Tridecanoic acid, 4,8,12-trimethyl, methyl ester	ND	0.22	0.18	ND
Cis-10-Heptadecenoic acid	ND	0.67		ND
Linoleic acid ethyl ester	ND	12.28	ND	ND
Methyl 5,11,14-eicosatrienoate	ND	ND	1.35	ND
Pentadecanoic, methyl ester (C15:0)	ND	ND	ND	1.35
7,10-Hexadecadienoic acid, methyl ester	ND	ND	ND	0.28
Heneicosanoic acid, methyl ester	ND	ND	ND	1.6
SFA	26.25	25.80	27.62	38.28
MUFA	17.38	5.68	23.74	36.24
PUFA	19.33	43.71	39.44	3.14

Analysis of Fatty acid profile based on the methyl ester of fatty acids (Area% of fatty acids) of *Nostoc spongiaeforme* grown under different temperature.

ND: Not detected; SFA: saturated fatty acid; MUFA: mono-unsaturated fatty acid; PUFA: poly-unsaturated fatty acid

**Table 2:** Analysis of Fatty acid profile based on the methyl ester of fatty acids (Area% of fatty acids) of *Nostoc calcicola* (Marine cyanobacteria) grown under different temperature.

Fatty acids	Percent of fatty acids (% of total fatty acids)			
	10°C	20°C	30°C	40°C
9-Hexadecenoic acid, methyl ester (C16:1)	6.19	7.05	5.95	2.44
Hexadecenoic acid, methyl ester (C16:0)	23.66	28.92	26.28	21.77
9,12-Octadecadienoic acid, methyl ester (C18:2)	13.62	10.62	17.63	17.91
9,12,15-Octadecatrienoic acid, methyl ester	27.51	22.4	20.73	ND
9-Octadecenoic acid methyl ester (C18:1)	10.68	2.53	7.38	28.29
Octadecanoic acid (C18:0)	5.21	11.57	4.7	7.84
Glycidyl palmitate	0.33	0.3	0.27	0.65
8,11,14-Docosatrienoic acid, methyl ester	0.41	0.36	ND	ND
Methyl tetradecanoate (C14:0)	ND	0.34	0.27	ND
Cis-10-Heptadecenoic acid	ND	0.4	0.46	ND
Octadecanoic acid, 3-hydroxy-methyl ester	ND	0.31	ND	ND
Cis-10-Nonadecenoic acid, methyl ester (C19:1)	ND	ND	0.69	ND
2,6,10-Trimethyltridecane	ND	ND	ND	0.57
Cyclopropaneoctanoic acid, 2-octyl-methyl ester	ND	ND	ND	0.51
SFA	29.20	41.44	31.52	30.77
MUFA	16.87	9.98	14.48	30.73
PUFA	41.54	33.46	38.36	17.91

## DISCUSSION

The effect of growth temperature on the fatty acid composition of two *Nostoc* species is reported through our present study. The result of fatty acid composition

indicated that both the *Nostoc* species displayed variation in the saturated and unsaturated fatty acid (MUFA & PUFA) to varying growth temperature. Through our result, it could be seen that in both *Nostoc* species, the saturated fatty acid decreased at low

temperature, whereas in the same condition unsaturated fatty acid (MUFA and PUFA) increased. Due to this, the degree of unsaturated fatty acid in both the *Nostoc* species increased. However, *N. calcicola* showed higher poly-unsaturated fatty acids as compared to control. It could be due to this reason *N. calcicola* showed a better growth rate compared to *N. spongiaeforme* (data not shown). As high degree of unsaturated may help the cells to maintain the membrane fluidity under cold conditions. Seto et al., (1984) reported an increase in the polyunsaturated fatty acid at low temperature in *Chlorella* which could be due to the sensitivity of the fatty acid biosynthetic enzymes. A similar pattern was observed by Chen et al. (2008) who reported an increase in the polyunsaturated fatty acid in *Nitzschia laevis* at low temperature. Our study was also on par with Jiang and Chen, 2000, their study also showed an increase in the polyunsaturated fatty acid at low temperature in *Cryptocodinium cohnii* a marine microalga. On the other hand, when the temperature was increased to 40°C there was a drastic reduction in the degree of unsaturated fatty acid. The decrease in the unsaturated fatty acid was escorted by the enhancement of saturated fatty acid. This may suggest that both *Nostoc* species was able to survive at higher growth temperature by adapting themselves to producing more saturated fatty acid compared to low temperature (Jiang and Chen, 2000). Through our result, it was also seen that, both the *Nostoc* species produced a maximal degree of polyunsaturated fatty acid when grown at 20°C and 30°C (control).

## CONCLUSION

Our study concludes that the varying growth temperature showed effects on the fatty acid composition of *N. spongiaeforme* and *N. calcicola*. The degree of fatty acid saturation increased at higher growth temperature which may have helped the *Nostoc* species to withstand the extreme temperature stress. Whereas, in order to maintain the fluidity of the membrane, on the other hand, low temperature resulted in the enhancement of unsaturated fatty acid. The temperature between 20°C to 30°C showed maximum fatty acid production in both the *Nostoc* species. Hence,

20°C-30°C is recommended temperature for producing maximum amount of fatty acid content in mesophilic nitrogen fixing cyanobacteria for biofuel production.

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## CONFLICTS OF INTEREST

The authors declare that they have no potential conflicts of interest.

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