

ORIGINAL ARTICLE

**Heavy metal accumulation potential and medicinal property of  
*Bacopa monnieri*- a paradox.**

**Hussain K<sup>1\*</sup> Abdussalam AK<sup>2</sup>, Ratheesh Chandra P<sup>2</sup>, Nabeesa Salim<sup>2</sup>**

<sup>1</sup> Dept. of Botany, Unity Women's College, Manjeri. Malppuram. Kerala-676122, India

<sup>2</sup> Division of Plant Physiology and Biochemistry, Dept.of Botany, Calicut University, Kerala-673635, India

Phone: + 9895501751

E-mail: [hussainkoorimannil@gmail.com](mailto:hussainkoorimannil@gmail.com)

Received August 13, 2011

Bioaccumulation of Mercury and Cadmium in *Bacopa monnieri* (L.) Pennell, cultivated in Hoagland medium artificially contaminated with micro quantities of HgCl<sub>2</sub> and CdCl<sub>2</sub> is investigated. Bioaccumulation potential of *B. monnieri* is more towards Cd than Hg. Absorption and translocation of Hg and Cd are proportional to the availability of the metal in the growth media and period of growth. Effect of acidic pH showed enhanced accumulation while basic pH resulted in significant reduction in the accumulation of Hg and exorbitant reduction of Cd. As a result of combined treatment of HgCl<sub>2</sub> and CdCl<sub>2</sub>, accumulation was very low in both acidic and basic pH. In addition to pH, antagonistic effect of Ca<sup>2+</sup> present in lime water which was added to raise the pH of the growth medium also controls the accumulation and absorption of Hg and Cd ions. Bioaccumulation of Hg and Cd in *B. monnieri* reveals the phytoremediation potential while the bioaccumulation is hazards to health since the plant is highly medicinal and one important ingredient of many Ayurvedic preparations.

*Key words: Bioaccumulation, Bacopa monnieri, Phytoremediation*

## ORIGINAL ARTICLE

## Heavy metal accumulation potential and medicinal property of *Bacopa monnieri*- a paradox.

Hussain K<sup>1\*</sup> Abdussalam AK<sup>2</sup>, Ratheesh Chandra P<sup>2</sup>, Nabeesa Salim<sup>2</sup>

<sup>1</sup> Dept. of Botany, Unity Women's College, Manjeri. Malppuram. Kerala-676122, India

<sup>2</sup> Division of Plant Physiology and Biochemistry, Dept.of Botany, Calicut University, Kerala-673635, India

Phone: + 9895501751

E-mail: [hussainkoorimannil@gmail.com](mailto:hussainkoorimannil@gmail.com)

Received August 13, 2011

Bioaccumulation of Mercury and Cadmium in *Bacopa monnieri* (L.) Pennell, cultivated in Hoagland medium artificially contaminated with micro quantities of HgCl<sub>2</sub> and CdCl<sub>2</sub> is investigated. Bioaccumulation potential of *B. monnieri* is more towards Cd than Hg. Absorption and translocation of Hg and Cd are proportional to the availability of the metal in the growth media and period of growth. Effect of acidic pH showed enhanced accumulation while basic pH resulted in significant reduction in the accumulation of Hg and exorbitant reduction of Cd. As a result of combined treatment of HgCl<sub>2</sub> and CdCl<sub>2</sub>, accumulation was very low in both acidic and basic pH. In addition to pH, antagonistic effect of Ca<sup>2+</sup> present in lime water which was added to raise the pH of the growth medium also controls the accumulation and absorption of Hg and Cd ions. Bioaccumulation of Hg and Cd in *B. monnieri* reveals the phytoremediation potential while the bioaccumulation is hazards to health since the plant is highly medicinal and one important ingredient of many Ayurvedic preparations.

*Key words: Bioaccumulation, Bacopa monnieri, Phytoremediation*

*Bacopa monnieri* (L.) Pennell is a fast growing wetland species and is well adapted to thrive in polluted areas which receive regular flushing of sewage waste and industrial effluents. Lenka *et. al.*, (1992) reported that *B. monnieri* accumulate mercury in the roots and shoots of plants growing in the wetlands in the vicinity of Chloralkali industry.

This plant has been recommended as an agent for phytoremediation (Sinha and Chandra, 1990, Sinha *et. al.*, 1996, Sinha, 1999, Yadav *et. al.* 2005). Potential of *B. monnieri* to accumulate As, Cd, Cr, Cu, Fe, Hg, Mn, Ni, Pb and Zn has been reported by analyzing the quantities of these elements in the

naturally growing plants collected from different polluted areas of Kerala (Hussain *et. al.* 2010).

Mercury is a global pollutant that cycle through bacteria, air, soil and water as a result of natural and anthropogenic activities and hence plants are directly exposed to this toxic heavy metal. Even though angiosperms have not yet been reported as tolerant to mercury (Lepp, 1981, Orcutt and Nilson, 2000; Cseh, 2002) and no plant has been identified as mercury hyper-accumulator (Henry, 2000, Raskin and Ensely, 2000), considerable absorption and accumulation of mercury have been reported in *B. monneri* (Sinha and Chandra, 1990, Sinha *et. al.* 1996), *Brassica jencea* (Moreno *et. al.* 2005a, b, 2008). *Chromolaena odorata* is an accumulator of Hg and has been recommended as a phytoremediant (Velasco-Alinsug *et al.*, 2005).

Cadmium is a wide spread toxic heavy metal of geogenic and anthropogenic pollutant (Prasad, 1997; Salt *et. al.* 1998; Orcutt and Nilson, 2000. Perfus-Barbeoch *et. al.* 2002) Phytoremediation of cadmium has been effectively performed by *Silene vulgaris* (Joop *et. al.* 1994), *Brassica juncea* (David *et.al.* 1995, Ishikawa *et.al.* 2006, *Polygonum thumberhii* (Shinmachi *et. al.*, 2003) *Potomegoton pectinatus* (Rai *et. al.* 2003) *Salia viminalies* (Mleezek *et. al.*, 2009).

*Bacopa monneri* has been used as important ingredient of Ayurvedic system of medicine for centuries. Traditionally the entire plant is used as a brain tonic to enhance memory, development, learning ability, concentration etc. and the compounds responsible for pharmacological effect include alkaloids, saponins and sterols (Nair, 1987, Wohlmuth, 2001, Anonymous, 2004).

The medicinal use and heavy metal bioaccumulation potential of *B. monneri* are found to be paradoxical because consumption of

Ayurvedic medicines containing this species which accumulates considerable quantities of toxic metals may lead to serious health hazards. So the present study was undertaken to assess the potential of *Bacopa monneri* to absorb and accumulate mercury and cadmium by growing the plant in Hoagland nutrient solution artificially contaminated with different concentration of HgCl<sub>2</sub> and CdCl<sub>2</sub>. Since pH of the nutrient solution is found to change the toxic effect of Mercury and Cadmium in *B. monneri* (Hussain, 2007) effect of different ranges of pH on bioaccumulation potential was another important objective of this study.

## MATERIAL AND METHODS

*Bacopa monneri*(L.) Pennell cuttings of 7-8 cms consisting of 5-6 nodes were collected from the medicinal plant collection of Calicut University Botanical garden. Rooted propagules were planted in Hoagland solution prepared following the modified method of Epstein (1972) as described by Taiz and Zeiger (1991). Three sets of experiments were conducted to assess the bioaccumulation pattern and potential of *B. monneri* towards mercury and cadmium.

### Dose response studies by mercuric chloride and cadmium chloride

For the treatments with HgCl<sub>2</sub> nutrient solution was contaminated with three concentrations of HgCl<sub>2</sub> (2, 5 and 10 µM) and based on dose- response studies conducted, about 50% growth retardation and reproducible responses were observed in 10 µM HgCl<sub>2</sub>. Similarly, three concentrations (10, 20 and 30 µM) solutions of CdCl<sub>2</sub> proved similar responses in 30 µM concentration. So, 10 and 30 µM concentrations of HgCl<sub>2</sub> and CdCl<sub>2</sub> respectively were selected for the experiments. Plants cultivated in the Hoagland nutrient medium served as the control.

Good quality plastic containers of 12x8.5x3 cm were used for nutrient culture. Plastic wire of 1mm diameter was tied on the mouth of the container interweaving length-wise and breadth-wise forming a mesh to provide mechanical support to the propagules. Hoagland medium with known quantities of HgCl<sub>2</sub> and CdCl<sub>2</sub> were within repaired contaminating rooted propagules were planted in the nutrient medium by inserting through the mesh without disturbing the root system. The gap provided underneath facilitated room for growth of

root system. All experimental containers were kept under green house condition.

Random sampling was followed to collect plants from ten replicate containers of each treatment and control at an interval of 2 days up to 12 days. Propagules collected were cut in to root, stem and leaf and were used for bioaccumulation studies of Hg and Cd.

#### Effect of additional supply of HgCl<sub>2</sub> and CdCl<sub>2</sub>:

To study the effect of additional Hg and Cd content on growth for prolonged period, the schedule of treatments is given below (Table 1):

**Table 1.** Quantitative schedule of Hg and Cd treatments

Treatment	Concentration n	Interval - Days			
		20	30	40	50
HgCl <sub>2</sub>	10 µM	Quantity of Hg/Cd µg per container			
		500	600	700	800
CdCl <sub>2</sub>	30 µM	840	1008	1176	1344

#### Effect of pH on the accumulation of Mercury and Cadmium

##### Acidic pH

Distilled water was used as the growth medium because change in pH resulted in the precipitation of nutrients in the Hoagland solution pH was adjusted to 5.5 by adding known quantities of 1M NH<sub>4</sub>Cl solution, to distilled water and solutions of HgCl<sub>2</sub> and CdCl<sub>2</sub>, were added to the containers to obtain the concentrations 20 and 30 µM respectively.

##### Alkaline pH

To distilled water, known quantity of calcium hydroxide solution was added to obtain alkaline pH (7.5). HgCl<sub>2</sub>/CdCl<sub>2</sub> solution was added to get 10 and 30µM respectively. Concentrations of Hg Hg<sup>2+</sup> and

Cd<sup>2+</sup> added to growth medium under different pH are given below (Table 2).

#### Estimation of Mercury and Cadmium

Plant parts-root, stem and leaf tissues of *B. monnieri* were sampled for analyzing Mercury and Cadmium contents. Samples were prepared according to the method of Allan (1969). Known weight of the samples were wet digested by refluxing in 10 ml of nitric acid, and perchloric acid in the ratio of 10: 4 until the solution became colorless by using Kjeldhahls flasks heated in a sand bath and digest was analyzed to estimate mercury and cadmium by using Atomic Absorption Spectrophotometer (Perkin Elmer model Analyst 300). All experiments were repeated a minimum of five times.

**Table 2.** Quantitative treatments at different pH

Medium	pH	Treatments and quantity given		
Distilled water	6.8	HgCl <sub>2</sub> (400µgHg)	CdCl <sub>2</sub> (224µgCd)	HgCl <sub>2</sub> +CdCl <sub>2</sub> , (400µgHg+224µgCd)
Acidic medium	5.5	do	do	do
Basic medium	7.5	do	do	do
Hoagland	6.2	do	do	do

## RESULTS

During growth for 12 days, accumulation of mercury was maximum in roots followed by stem and leaves during all intervals (Fig. 1) Mercury accumulated in the root tissue remained unchanged during growth compared to the stem and leaf tissue which showed enhanced rate of accumulation.

Accumulation of cadmium also was maximum in root tissue and order of accumulation was Root>Stem>Leaf. Cadmium content accumulated in the root was progressively increased during growth. Similar trend was seen in stem tissue and Cd content of leaf was very low. (Fig. 2)

When the plants were exposed to additional doses 100µg each of Hg at an interval of 10 days each during 50 days, Hg concentration per plant was increased Cd proportional to the content given (Table 4). The quantity accumulated at each interval was approximately 25% of the total given in the medium. In the case of cadmium also more or less similar pattern of accumulation was observed (Table 4). But the quantity accumulated was above 50% of the total given in the medium at each interval.

Effect of acidic (5.5) - and alkaline (7.5) pH showed remarkable differences in metal accumulation (Table 4). At acidic pH, mercury accumulation was more than the control and showed two fold increase compared to the alkaline medium. Propagules grown in acidic medium accumulated Cd content more than 8 times in comparison with the control whereas in alkaline medium, only very low quantity was accumulated (less than 1%). When mercury treatment was given in Hoagland nutrient medium (pH 6.0) accumulation was similar to that of the control and acidic pH whereas Cd content was significantly reduced compared to that of the acidic pH but was more than the control (Table 3). Combined treatment of HgCl<sub>2</sub> and CdCl<sub>2</sub> showed very low amount of Hg in all treatments and control whereas Cd content was reduced compared to their individual treatment. Similarly in alkaline and acidic media, Hg accumulation showed negligible difference whereas Cd accumulation in the acid medium remained unchanged, but in alkaline medium an exorbitant reduction was observed compared to their respective control. (Table 3).

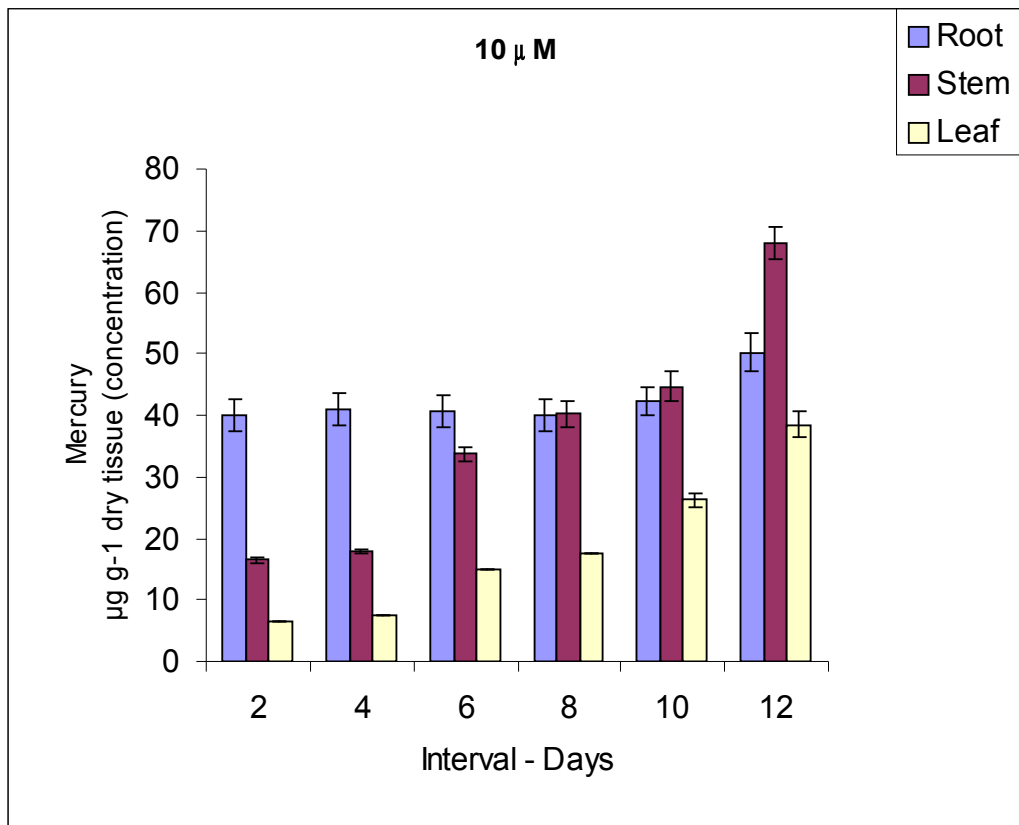
**Table 3** Bioaccumulation of Mercury and Cadmium in *Bacopa monneri* during 50 days of growth in different concentrations

Treatment	Concentration	Interval (days)	Quantity given	Accumulation mg
HgCl <sub>2</sub>	10µM	20	500	121± 4.6 (24.2)
		30	600	164± 4.1 (27.3)
		40	700	189± 5.2 (27.3)
		50	800	204± 5.7 (25.5)
CdCl <sub>2</sub>	30 µM	20	840	409± 12.8 (48.6)
		30	1008	491± 11.6 (48.7)
		40	1176	508± 12.4 (50.3)
		50	1344	565± 12.8 (49.9)

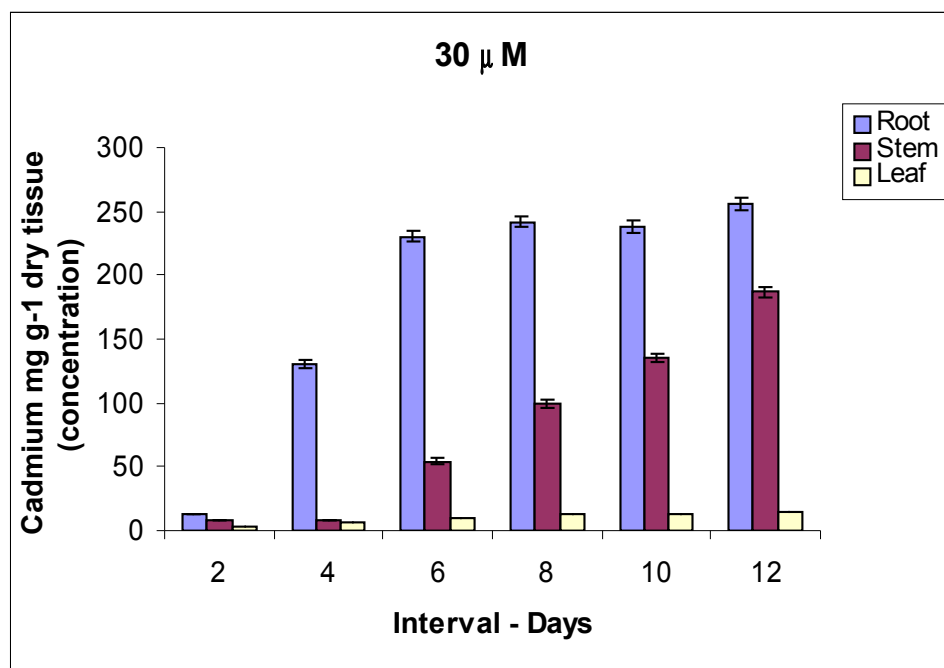
Values in parenthesis are percentage of accumulation

**Table 4** Effect of pH on mercury and cadmium uptake in *B. monneri* during growth in different media (mg<sup>-1</sup> dry tissue)

Treatments		Distilled water pH 6.8	Acidic pH 5.0	Basic pH7.5	Hoagland pH 6.2
HgCl <sub>2</sub> 400µgHg	Hg	54.00±1.03	68.88±.1.04	36.20±0.82	66.3±1.04
CdCl <sub>2</sub> 224µgCd	Cd	35.46±0.90	266.80±2.07	2.39±0.03	72.30±1.04
HgCl <sub>2</sub> + CdCl <sub>2</sub> 400µgHg+ +224µgCd)	Hg	0.60±0.01	0.47±0.01	0.77±0.01	0.60±0.01
	Cd	23.98±0.82	28.80±1.02	1.02±0.02	1.90±0.01



**Figure 1** Accumulation of Mercury contents in different parts of *Bacopa monnieri* treated with  $\text{HgCl}_2$  during growth.  $\mu\text{g/g}^{-1}$  dry tissue (concentration)



**Figure 2** Accumulation of Cadmium contents in different parts of *Bacopa monnieri* treated with  $\text{CdCl}_2$  during growth.  $\text{mg/g}^{-1}$  dry tissue (concentration)

## DISCUSSION

During 12 days of growth under the simulated laboratory conditions, mercury content of root was almost uniform whereas stem and leaves showed significant ( $p < 0.01$ ) increase (Fig.1). The mercury uptake pattern of *Bacopa monnieri* is similar to that reported in *Chromolaena odorata* which has been recommended as a phytoremediant. (Velasco-Alinsug *et.al*, 2005). About  $40\mu\text{g gram Hg g}^{-1}$  dry tissue of roots appear to be the thresholded level of accumulation to which the plant is tolerant and above this level, it may cause toxicity to root system as suggested by Bauford *et.al*, (1977). Notwithstanding, Additional application of  $\text{HgCl}_2$  and prolonged growth period exhibit proportionate increase in accumulation on per plant basis (Table 4) and so this observation indicates the phytoremediation potential of *B. monnieri* Although no plant has yet been identified as natural hyper-accumulator of Hg (Henry, 2000; Ruskin and Ensley, 2000), transgenic plants such as *Arabidopsis thaliana*, *Liriodendron tulipifera* and *Nicotiana tabacum* are reported to be capable of converting methyl mercury to  $\text{Hg}^{2+}$  and are having the potential of phytoremediation in alleviating Hg polluted areas (Bizily *et. al.*, 1997; Rugh *et al.*, 1996; 1998). In the present study *Bacopa monnieri* is found to accumulate about 25% of the Hg content available in the growth medium. This is inconformity with views of (Sinha *et al.*, (1996) and Sinha (1999) who recommended *Bacopa monnieri* as a plant for phytoremediation of metals such as Cd, Cr, Cu, Hg, Mn and Pb from contaminated water and wetlands. Nevertheless, this plant is not a hyper accumulator of heavy metals.

Bio accumulation potential of *Bacopa monnieri* towards cadmium is not similar to mercury on per plant basis. About 50% Cd present in the medium is absorbed during prolonged period of growth (Table

4). Accumulation of Cadmium is also maximal in roots and this metal reported to be easily taken up by the roots and transported to the leaves since  $\text{Cd}^{2+}$  ion are fast mobile in plants (Siedlecka and Krupa, 1997). According to Sersen *et. al.*, (2005), Maize plants grown in nutrient medium containing Cd is able to absorb Cd and the accumulation is proportional to the availability of the metal. The accumulation pattern of Cd in the roots of *Bacopa monnieri* is almost in consistent with the views of Sersen *et. al.*, (2005) because Cd accumulation is proportional to the increase of  $\text{CdCl}_2$  concentration in the nutrient medium. According to Sanita-di-Toppi and Gabbrielli (1999) immobilization of Cd by binding to the cell wall is one of the causes of Cd hyperaccumulation in plants. Many plants such as *Potamogeton pectinatus* (Rai *et.al.*, 2003) *Arabidopsis thaliana* (Perfus –Barbeoch *et. al.*, 2002. *Phragmites australis* (Ederli *et.al*, 2004), *Cannabis sativa* (Linger *et. al.*, 2005) *Brassica juncea* (Ishikawa *et. al*, 2006) are reported as hyper accumulators of Cd. Eventhough most of the Cd accumulators are recommended for phytoremediation (Pilon-Smits, 2005), translocation of  $\text{Cd}^{2+}$  to rice grains causing health hazards have been reported recently by Tanaka *et. al.*, (2007)

Accumulation of Hg and Cd by *Bacopa monnieri* is dependent on the presence of other ions as well as the pH of the growth medium. The plants cultivated in distilled water containing  $\text{HgCl}_2$  and  $\text{CdCl}_2$  having the pH 5.5 showed a slight increase in accumulation of Hg (Table 4.). The accumulation of Cd was 8 times higher. The behavior of *Bacopa monnieri* towards pH changes in the absorption mode of Cd is in accordance with the findings of Hattori *et. al.*, (2006) in *Helianthus* and *Hibiscus* where a doubling of Cd uptake was occurred by the application of  $\text{Cl}^-$  to reduce soil pH. Accumulation of Cd is reduced exorbitantly in basic pH in



comparison with the pH 5.5 (Table 4). This may not be due to the effect of pH alone. The control of influx and /or antagonistic effect of  $\text{Ca}^{2+}$  due to the addition of Calcium hydroxide to the medium cannot be ruled out in accordance with the view of Hough *et al.*, (2003) in wheat where liming to reduce soil pH resulted in reduced Cd uptake. According to Kim *et al.*, (2002)  $\text{Ca}^{2+}$  inhibits Cd accumulation in rice roots by substituting  $\text{Ca}^{2+}$  to  $\text{Cd}^{2+}$  ions. Perfus-Barbeoch *et al.*, (2002) suggested that in *Arabidopsis thaliana*,  $\text{CdCl}_2$  induced stomata closure by controlling  $\text{Ca}^{2+}$  channels of plasma membrane of guard cells and  $\text{Cd}^{2+}$  mimics  $\text{Ca}^{2+}$  channels and enter guard cell through calcium channels. According to Ried *et al.* (2003) Cd accumulation in plants is due to many physical similarities between  $\text{Ca}^{2+}$  and  $\text{Cd}^{2+}$  in such a way that the ionic radius of  $\text{Ca}^{2+}$  (9PM) and that of  $\text{Cd}^{2+}$  (7PM) causes sharing of  $\text{Ca}^{2+}$  channels with  $\text{Cd}^{2+}$  for the absorption and translocation in potato tuber.

*Bacopa monneri* plants grow wildly in wetland which naturally get contaminated with sewage and other waste water and considerable accumulation of Hg, Cd, Cr, Cm, Mu and Pb in naturally growing of *Bacopa monneri* plants have been reported (Hussain-Koorimannil *et al.* 2010). This stimulated experimental study confirms the accumulation potential of this plant. Collection of *B. monneri* plants from contaminated wetlands is a common practice of manufacturers of Ayurvedic medicines and food supplements and consumption of these products by human beings will lead to serious health hazards.

The effect of changing pH on the combination of  $\text{HgCl}_2$  and  $\text{CdCl}_2$  treatment reveals significant inhibition of Hg uptake and partial control over  $\text{CdCl}_2$ . The antagonistic effect Hg and Cd on one hand and that of  $\text{Ca}^{2+}$  and  $\text{NH}_4^+$  on the other may be the plausible reason. In *Bacopa monneri* increased

pH due to the addition of calcium hydroxide to nutrient medium control the entry of  $\text{Cd}^{2+}$  through  $\text{Ca}^{2+}$  channels and the overwhelming flux of the latter may be due to the similar ionic radius of  $\text{Cd}^{2+}$  and  $\text{Ca}^{2+}$  as suggested by (Raid *et al.*, 2003). In this context, liming can be recommended as a measure against  $\text{Cd}^{2+}$  and  $\text{Hg}^{2+}$  absorption and translocation. On the contrary, reduced pH of the growth medium (5.5) by adding  $\text{NH}_4\text{Cl}$  (chlorination) results in signified increase of Hg and Cd uptake (Table 4).

## CONCLUSION

The medicinal property and wide use of *Bacopa monneri* as an ingredient of many Ayurvedic medicines and food supplements on one hand and the bioaccumulation potential and phytoremediation efficacy on the other are paradoxical. The absorption and accumulation of Hg and Cd is maximum at acidic pH and very low at alkaline pH by *B. monneri* convey that, for medicinal and phytoremediation purposes, cultivation in alkaline soil/water and acidic soil/water respectively is recommended.

## REFERENCES

- Allan, J.E. (1969), *The preparation of agricultural samples for analysis by Atomic Absorption Spectrometry*. S.I.S. Edition; *Varian Techtron Bulletin*. 12-69.
- Anonymous. (2004), *Bacopa monniera* – monograph. *Altern. Med. Rev.* **9**, 79-85.
- Beauford, W., Barber, J. and Barringer, A.R. (1977), Uptake and distribution of mercury within higher plants. *Physiol. Plant.* **39**, 261-265.
- Bizily, S., Rugh, C., Summers, A.O. and Meagher, R.B. (1997), Phytoremediation of methyl mercury pollution: Mer B expression in *Arabidopsis thaliana* confers resistance to organo mercurials. *Proc. Nalt. Acad. Sci. USA.* **96**, 6808-6813.

- Cseh, E. (2002), Metal permeability, transport and efflux in plants. In: M.N.V. Prasad and K. Strzalka (Eds.). *Physiology and Biochemistry of Metal Toxicity and Tolerance in Plants*. 1-36.
- David, E.S., Rogfer, C.P., Ingrid, J.P. and Ilya, R. (1995), Mechanisms of cadmium mobility and accumulation in Indian mustard. *Plant Physiol.* **109**, 1427-1433.
- Ederli, L., Reale, L., Ferrauti, F. and Pasqualini, S. (2004), Responses induced by high concentration of cadmium in *Phragmites australis* roots. *Physiol. Plant.* **121**, 66-74.
- Epstein, E. (1972), *Mineral Nutrition of Plants, Principles and Perspectives*. John Wiley & Sons, New York.
- Hattori, H., Kuniyasu, K., Chiba, K. and Chino, M. (2006), Effect of chloride application and low soil pH on cadmium uptake from soil by plants. *Soil Sci. Plant Nutr.* **52**, 89-94.
- Henry, J.R. (2000), Phytoremediation of mercury. In: *An Overview of the Phytoremediation of Lead and Mercury: A Report for the U.S. Environ. Protec. Agency* Washington, D.C. USA. 44-46.
- Hoagland, D.R. and Arnon, D.I. (1950), The water culture method of growing plants without soil. *Colif. Agric. Expal. Stn. Circ.* **347**, 461-465.
- Hough, R.L., Hough, R.L., Young, E.D. and Crout, N.M.J. (2003), Modelling of Cd, Cu, Ni, Pb and Zn uptake by winter wheat and forage maize, from a sewage disposal farm. *Soil Use Manage.* **19**, 19-27.
- Hussain, K. (2007), Ecophysiological aspects of *Bacopa monnieri* (L.) Pennell. Thesis submitted to the University of Calicut, Kerala.
- Hussain-Koorimannil., Abdussalam, A.K., Ratheesh-Chandra, P. and Nabeesa Salim. (2010), Bio accumulation of heavy metals in *Bacopa monnieri* (L.) Pennell growing under different habitat. *Int. J. Ecol. & Develop.* **15**, 66-73.
- Ishikawa, S., Ae, N., Murakami, M. and Wagatsuma, T. (2006), Is *Brassica juncea* a suitable plant for phytoremediation of cadmium in soils with moderately low cadmium contamination? – Possibility of using other plant species for Cd-phytoextraction. *Soil Sci. Plant Nutr.* **52**, 32-42.
- Joop, A.K., Marjolein, D., Paul, L.M.K., Henk, S., Jos, A.C.V. and Wilfried, H.O.E. (1994), Phytochelatin in cadmium sensitive and cadmium tolerant *Silene vulgaris*. *Plant Physiol.* **104**, 255-261.
- Kim, Y.Y., Yang, Y.Y. and Lee, Y. (2002), Pb and Cd uptake in rice roots. *Physiol. Plant.* **116**, 368-372.
- Lenka, M., Panda, K.K. and Panda, B.B. (1992), Monitoring and assessment of mercury pollution in the vicinity of a chloralkali plant, IV: Bioconcentration of mercury *in situ* aquatic and terrestrial plants at Ganjam, India. *Arch. Environ. Contam. Toxicol.*, **22**, 195-202.
- Lepp, N.W. (1981), *Effect of Heavy Metal Pollution on Plants*. Vol. 2. Applied Science Publishers, London.
- Linger, P., Ostwald, A. and Haensler, J. (2005), *Cannabis sativa* L., growing on heavy metal contaminated soil: Growth, cadmium uptake and photosynthesis. *Biol. Plant.* **49**, 567-576.
- Mleezek, M., Kasyewski, L.M., Kaezmarekm, Z.M., Rissmann, I. and Gabinski, P. (2009), Efficiency of selected heavy metals accumulation by Saliva roots *Environ. Exp. Bot.* **65**, 48-53

- Moreno, F.N., Anderson, C.W.N., Stewart, R.B. and Robinson, B.H. (2008), Phytofiltration of mercury –contaminated water: Volatilisation and plant –accumulation aspects. *Environ. Exp. Bot.* **62**, 78-85.
- Moreno, F.N., Anderson, C.W.N. Stewart, R.B., Robinosn, B.H., Nomura, R., Ghomshei, M. and Meech, J.A. (2005b), Effect of thioglands on plant –Hg accumulation and volatilisation from mercury contaminated mine tailings. *Plant soil*, **275**, 233-246.
- Nair, K.K.N. (1987), Medhya Rasayana Drug 'Brahmi' - Its Botany, Chemistry and Uses. *J. Econ. Tax. Bot.* **11**, 359-365.
- Orcutt, D.M. and Nilsen, E.T. (2000), *Physiology of Plants Under Stress: Soil and Biotic Factors*. John Wiley & Sons, Inc. New York.
- Perfus-Barbeoch, L., Leonhardt, N., Vavasseur, A. and Forestier, C. (2005), Heavy metal toxicity: Cadmium permeates through calcium channels and disturbs the plant water status. *The Plant J* 2002; **32**, 539-548.
- Pilon-Smits, E. (2005), Phytoremediation. *Annu. Rev. Plant. Biol.* **56**, 15-39.
- Prasad, M.N.V. (1997), Trace metals In: M.N.V. Prasad (Ed.). *Plant Ecophysiology*. John Wiley & Sons, Inc. 207-249.
- Rai, U.N., Tripathi, R.D., Vajpayee, P., Pandey, N., Ali, M.B. and Gupta, D.K. (2003), Cadmium accumulation and its phytotoxicity in *Potamogeton pectinatus* L. (Potamogetonaceae). *Bull. Environ. Contam. Toxicol.* **70**, 566-575.
- Raskin, I. and Ensley, B.D. (2000), *Phytomediation of Toxic Metals Using Plants to Clean up the Environment*. John Wiley & Sons. Inc. New York.
- Reid, R.J., Dunbar, K.R. and McLaughlin, M.J. (2003), Cadmium loading into potato tubers: the roles of the periderm, xylem and phloem. *Plant Cell Environ.* **26**, 201-206.
- Rugh, C.L., Senecoff, J.F., Meagher, R.B. and Merkle, S.A. (1998), Development of transgenic yellow poplar for Hg phytoremediation. *Natur. Bio. Technol.* **16**, 925-928.
- Rugh, C.L., Wilde, H.D., Stack, N.M., Martin-Thompson, D., Summers, A.O. and Meagher, R.B. (1996), Mercuric ion reductase and resistance in transgenic *Arabidopsis thaliana* plants expressing a modified bacterial mer A gene. *Proc. Natl. Acad. Sci. USA.* **93**, 3182-3187.
- Salt, D.E., Smith, R.D. and Raskin, I. (1998), Phytoremediation. *Annu. Rev. Plant Physiol. Mol. Biol.* **49**, 643-668.
- Sersen, F., Clik, G., Havranek, E. and Sykorova, M. (2005), Bio-remediation by natural zeolite in plants cultivated in a heavy metal-contaminated medium. *Fresenius Environ. Bull.* **14**, 13-17.
- Shinmachi, F., Kumanda, Y., Noguchi, A. and Hasegawa, I. (2003), Translocation and accumulation of cadmium in cadmium tolerant *Polygonum thunbergii*. *Soil Sci. Plant Nutr.* **49**, 355-361.
- Siedlecka, A. and Krupa, Z. (1997), Cd/Fe interaction in higher plants – its consequences for the photosynthetic apparatus. *Photosynthetica* **36**, 321-331.
- Sinha, S. (1999), Accumulation of Cu, Cd, Cr, Mn, and Pb from artificially contaminated soil by *Bacopa monnieri*. *Environ. Monit. Assess.* **57**, 253-264.
- Sinha, S. and Chandra, P. (1990), Removal of Cu

- and Cd from water by *Bacopa monnieri* (L.). *Water Air Soil Pollut.* **51**, 271-276.
- Sinha, S., Gupta, M. and Chandra, P. (1996), Bioaccumulation and biochemical effect of mercury in the plant *Bacopa monnieri* (L.). *Environ. Toxicol. Wat. Qual.* **11**, 105-112.
- Taiz, L. and Zeiger, E. (2002), *Plant Physiology*: IIIrd edition, Sinauer Associates, Inc., Publishers, Sunderland, Massachusetts.
- Tanaka, K., Fujimaki, S., Fujiwara, T., Yoneyama, T. and Hayashi, H. (2007), Quantitative estimation of the contribution of the phloem in cadmium transport to grains in rice plants (*Oryza sativa* L.). *Soil Sci. Plant Nutr.* **53**, 72-77.
- Velasco-Alinsug, M.P., Rivero, G.C. and Quibuyen, T.A.O. (2005), Isolation of mercury-binding peptides in vegetative parts of *Chromolaena odorata*. *Z. Naturforsch.* **60c**, 252-259.
- Wohlmuth, H. (2001), Brahmi update. In: *Botanical Pathways, Information and Research on Botanical Medicine*, 8: 1. ([www.netresources.com.au/health/brahmi.pdt](http://www.netresources.com.au/health/brahmi.pdt)).
- Yadav, S., Sukla, O.P. and Rai, U.N. (2005), Chromium pollution and bioremediation. *Environ. News Archiv.* **11**, 1-4.